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# Ancient DNA reveals interstadials as a driver of the common vole population dynamics during the last glacial period

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- 5 Running title: Evolutionary history of common vole
- 6
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#### 48 1 Abstract

#### 49 Aim

- 50 The common vole is a temperate rodent widespread across Europe. It was also one of the most
- 51 abundant small mammal species throughout the Late Pleistocene. Phylogeographic studies of
- 52 its extant populations suggested the Last Glacial Maximum (LGM, 26.5–19 ka ago) as one of
- 53 the main drivers of the species' population dynamics. However, analyses based solely on extant
- 54 genetic diversity may not recover the full complexity of past population history. The main aim
- 55 of this study was to investigate the evolutionary history and identify the main drivers of the
- 56 common vole population dynamics during the Late Pleistocene.
- 57 Location
- 58 Europe
- 59 Taxon
- 60 Common vole (*Microtus arvalis*)
- 61 Methods

We generated a dataset comprising 4.2 kb-long fragment of mitochondrial DNA from 148 ancient and 51 modern specimens sampled from multiple localities across Europe and covering the last 60 thousand years (ka). We used Bayesian inference to reconstruct their phylogenetic

- 65 relationships and to estimate the age of specimens that were not directly dated.
- 66 Results

We estimate the time to the most recent common ancestor of all Last Glacial and extant common vole lineages to 90 ka ago and the divergence of the main mtDNA lineages present in extant populations to between 55 and 40 ka ago, earlier than previous estimates. We find multiple lineage turnovers in Europe in the period of high climate variability at the end of Marine Isotope Stage 3 (MIS 3; 57–29 ka ago) in addition to those found previously around the Pleistocene/Holocene transition. Conversely, data from the Western Carpathians suggest continuity throughout the LGM even at high latitudes.

74 Main conclusions

75 Our results suggest that the main factor affecting the common vole populations during the last

76 glacial period was the reduction of open habitats during the interstadial periods while the

- climate deterioration during the LGM had little impact on species' population dynamics.
- 78

Keywords: ancient DNA, *Microtus arvalis*, habitat, Late Pleistocene, paleoclimate, small
 mammals

81

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103

#### 104 **3 Introduction**

105The climatic and environmental changes during the last glacial period (ca. 115–11.7 ka106ago) had a great impact on the evolutionary histories of most species. It has been suggested that

107 species have responded to those changes according to their individual characteristics and there 108 is no basis for considering them as communities responding to climate and environmental 109 changes in the same manner (Baca et al., 2017; Lorenzen et al., 2011; Stewart, Lister, Barnes, 110 & Dalén, 2010). However, within the ecosystem, species depend on a whole range of 111 interactions at different trophic levels and across different ecological niches (Walther, 2010). 112 Thus, investigations of species with different adaptations can reveal the spectrum of responses 113 to the same climatic and environmental fluctuations and allow identification of the key factors 114 driving ecosystem responses (Cooper et al., 2015). Small mammals may be especially well 115 suited for such investigations as, in contrast to megafaunal species, they seem to be little 116 affected by activities of Palaeolithic hunter-gatherers and their population dynamics were 117 mainly driven by environmental changes.

118 The common vole *Microtus arvalis* (Pallas, 1779) is a temperate rodent species which 119 is present in most of continental Europe (Figure 1b). The species feeds primarily on leaves and 120 grasses and prefers well-drained grasslands, pastures, and alpine meadows from lowlands up to 121 ca. 3000 m above sea level. At present it mainly utilizes secondary habitats, such as agricultural 122 fields, where it is often considered a pest (Jacob, Manson, Barfknecht, & Fredricks, 2014). The 123 earliest remains of ancestral common voles (Microtus arvalis-group) in Europe date to ca. 0.6-124 0.5 Mya (Berto, Nadachowski, Pereswiet-Soltan, Lemanik, & Kot, 2021; Kučera, Suvova, & 125 Horáček, 2009; Maul & Markova, 2007). The fossil record from the last glacial period (115– 126 11.7 ka ago), attests to its continuous presence on most of the continent (Chaline, 1972; Horáček 127 & Ložek, 1988; Jánossy, 1986; Nadachowski, 1989). In many localities on the European Plain, 128 from France to Poland, the common vole was the most abundant small mammal, alongside the 129 collared lemming (Dicrostonyx torquatus) and the European narrow-headed vole 130 (Lasiopodomys anglicus) (Royer et al., 2016; Socha, 2014). The mitochondrial DNA (mtDNA) 131 phylogeography of extant populations of the common vole has been intensively studied, with 132 the aim of reconstructing the post-glacial history of the species. The extant mtDNA diversity is 133 partitioned into six divergent lineages with parapatric distribution: Western-South (WS), 134 Western-North (WN), Italian (ITA), Balkan (B), Central (CEN), and Eastern (E) (Bužan, 135 Förster, Searle, & Kryštufek, 2010; Haynes, Jaarola, & Searle, 2003; Heckel, Burri, Fink, 136 Desmet, & Excoffier, 2005; Stojak, McDevitt, Herman, Searle, & Wójcik, 2015) (Figure 1b). 137 Most of the previous studies estimated the time to the most recent common ancestor (tMRCA) 138 of the extant common vole populations to between 65 and 50 ka ago and subsequent divergence 139 of main lineages to between 50 and 20 ka ago (García et al., 2020; Heckel et al., 2005; Stojak 140 et al., 2016). In contrast, Fink et al. (2004) and Tougard et al. (2008) used fossil calibration to

141 suggest a much older diversification in the Middle Pleistocene. Analyses of nuclear DNA 142 revealed overall a very good correspondence with the spatial distributions of mtDNA lineages 143 (Fischer, Foll, Heckel, & Excoffier, 2014; Heckel et al., 2005) and detailed analyses of the 144 contact areas between lineages demonstrated admixture only in narrow hybrid zones (Beysard 145 & Heckel, 2014; Braaker & Heckel, 2009). However, the divergence time estimates of the 146 evolutionary lineages based on the nuclear data were generally much more recent than those 147 based on mtDNA and suggested that the diversification of common vole evolutionary lineages 148 took place during or after the LGM (Heckel et al., 2005; Lischer, Excoffier, & Heckel, 2014).

149 The present-day distribution of common vole mtDNA lineages (Figure 1b) was interpreted as evidence for the survival of common vole populations during the LGM in both 150 151 traditional Southern glacial peninsular refugia, as well as at higher latitudes, especially in 152 Central France (WN), north of the Alpine region (CEN) and in the Carpathian area (E) (Heckel 153 et al., 2005; Stojak et al., 2015; Tougard et al., 2008). Examination of nuclear DNA from 154 multiple European populations revealed a south-west cline of genetic diversity which was 155 interpreted as indicative of a westward expansion of the common vole at a time prior to the 156 LGM (Heckel et al., 2005), nevertheless the pre-LGM history of the species remains largely 157 unknown.

158 A detailed study of the Eastern mtDNA lineage suggested that it originated in the 159 Carpathian area and its current distribution is the result of an expansion which started after the 160 LGM with a possible bottleneck during the Younger Dryas (12.8–11.7 ka ago) (Stojak et al., 161 2016). Recently, an ancient DNA investigation of the common vole remains from the Western 162 Carpathians has shown the presence of the Eastern lineage from the early Holocene onwards. 163 However, it also showed that from at least 18 ka ago the Central lineage was present in this region suggesting a population replacement around the Pleistocene/Holocene transition (Baca 164 165 et al., 2020). This challenged the simple model of post-glacial recolonisation of the Eastern 166 lineage from the Carpathian refugium and suggested that the analyses based solely on extant 167 genetic diversity may not recover the full complexity of the Late Pleistocene population 168 dynamics. To refine the evolutionary history of common vole populations in Europe and to 169 compare it with the past histories of coeval cold-adapted species such as the collared lemming, 170 we generated and analysed a new mitochondrial dataset consisting of nearly 200 sequences 171 from Late Pleistocene and extant individuals.

#### **Material and Methods** 172 4

#### 173 4.1 **Ancient specimens**

174 Isolated lower first molars or mandible fragments with molars classified as M. arvalis 175 or Microtus sp. based on morphology of the occlusal surface were collected from various 176 palaeontological sites across Europe (Table S1). Each tooth was photographed at the Institute 177 of Systematics and Evolution of Animals, PAS.

#### 178 4.2 **Modern specimens**

179 DNA of modern specimens from various locations across Europe was extracted 180 previously (Table S2). A target 4.2 kb region of mtDNA spanning positions 12,000 to 16,247 181 according to the reference sequence (NC 038176; Folkertsma et al., 2018) was generated using 182 various approaches. It was either PCR amplified, sonicated, transformed into sequencing 183 libraries and sequenced or the genomic DNA was sonicated and transformed into sequencing 184 libraries. It was then either enriched for target fragment using in-solution hybridization and 185 sequenced or the target region was extracted from deep shotgun sequencing data (see Appendix 186 A1.1 and Table S2 for more details).

#### 187 4.3 Ancient DNA extraction, target enrichment and sequencing

188 DNA extraction and pre-PCR library preparation steps were performed in the dedicated ancient DNA laboratory at the Centre of New Technologies at the University of Warsaw. Each 189 190 tooth was thoroughly cleaned with ultra-pure water in a 2 ml tube and crushed with a pipette 191 tip. DNA was extracted following a silica spin column-based protocol optimized for retrieval 192 of short DNA molecules (Dabney et al., 2013). A negative control without biological material 193 was processed alongside each batch of 15 specimens. Double-stranded, double-indexed 194 sequencing libraries were produced from half of the DNA extract (20 µl) following a previously 195 established protocol (Meyer & Kircher, 2010) with minor modifications (Baca et al., 2019). For 196 some specimens that yielded low-quantity DNA additional double-indexed, single-stranded 197 sequencing libraries were prepared following the protocol proposed by Gansauge et al. (2020) 198 (see Appendix A1.3–A1.4 for more details).

199 Libraries were enriched for vole mitochondrial DNA using an in-solution hybridization 200 protocol described in Baca et al. (2019). Up to five libraries were pooled for hybridization 201 reaction. We performed two rounds of hybridization in 65°C for 22–24h each. After each round, 202 library pools were washed and amplified in triplicate for 10 to 15 cycles. Enriched library pools

were combined, quantified using qPCR and sequenced on Illumina NextSeq550 platform (MID
output, 2x75 bp kit; see Appendix A1.2–A1.5 and Tables S4–S5 for more details).

#### 205 4.4 Sequence processing

206 Sequencing reads were demultiplexed using bcl2fastq v. 2.19 (Illumina). Overlapping 207 reads were collapsed, adaptor and quality trimmed using AdapterRemoval v. 2.2.2 (Schubert, 208 Lindgreen, & Orlando, 2016). Then, reads were mapped to the common vole mtDNA genome 209 using the mem algorithm in bwa v. 0.7.17 (Li & Durbin, 2010). Duplicates, short (<30 bp) and 210 low mapping quality reads (mapq<30) were removed using samtools v. 1.9. Variants and consensus sequences were called using bcftools v. 1.9 (Li et al., 2009). Read alignments and 211 212 vcf files were inspected manually using Tablet v. 1.17 (Milne et al., 2013). Positions with 213 coverage below three were masked with N. If a base was supported by less than 75% of reads, 214 an IUPAC symbol was inserted. MapDamage v. 2.08 (Jónsson, Ginolhac, Schubert, Johnson, 215 & Orlando, 2013) was used to assess the damage patterns and length distribution of DNA 216 molecules. See Appendix A1.6 for more details.

## 217 4.5 Phylogenetic analyses and molecular dating of specimens

We used a Bayesian approach, implemented in BEAST 1.10.4 (Suchard et al., 2018), to estimate divergence times of common vole lineages and the age of specimens which were not directly dated. For the phylogenetic inference we used only ancient and modern sequences with at least 70% and 90% of the target mtDNA fragment (4.2 kb) recovered, respectively. Sequences were aligned with MAFFT v. 7.407 (Katoh & Standley, 2013). The best substitution model selected by jModelTest2 (Darriba, Taboada, Doallo, & Posada, 2012), TIM2+F+I+G4, was not easily available in BEAST we therefore used the closest available one: GTR +I+G.

225 First, we used Bayesian evaluation of temporal signal (BETS) (Duchene et al., 2020) to 226 check whether there is sufficient temporal resolution within our dataset to calibrate the 227 molecular clock. We used all directly radiocarbon dated (n = 20) and modern (n = 51) specimens 228 and tested four alternative models. In two of them, we assigned real sampling times to the 229 sequences (heterochronous analysis) and used either strict clock or uncorrelated relaxed 230 lognormal clock. In the two other models, we used the same sampling time for all sequences 231 (isochronous analysis) and applied either strict clock or uncorrelated relaxed lognormal clock 232 (see Appendix A1.7 for more details). Then, we performed the leave-one-out analysis on the 233 directly radiocarbon dated specimens to check the accuracy of the age estimates produced using 234 the available calibration dataset. In this analysis we estimated the age of each directly 235 radiocarbon dated specimen using all the remaining radiocarbon dated and modern specimens to calibrate the molecular clock. Next, we estimated the age of each ancient, not directly dated, specimen (n = 128) in a separate BEAST run, again using all directly dated and modern specimens to calibrate the molecular clock. Finally, we ran a joint analysis with all the sequences. For the specimens that were not directly dated we set a lognormal prior with a mean equal to the mean age estimated in the individual analysis and the range covering the 95% HPD interval of the individual estimate (see Appendix A1.7, Tables S6–S9 for more details).

The demographic reconstructions using the Bayesian Skyline and Bayesian SkyGrid methods implemented in BEAST 1.10.4 were performed for the WN lineage specimens from Spain (n=58). In the case of other localities either the number of available sequences was low or the assumption of population continuity (i.e., lack of lineage turnovers) were violated (see Appendix A1.8 for details).

## 247 4.6 Radiocarbon dating

Selected vole mandibles were pretreated for radiocarbon dating in the Department of 248 249 Human Evolution at the Max Planck Institute for Evolutionary Anthropology (MPI-EVA, Leipzig, Germany) following the protocol for <100 mg bone samples described in Fewlass et 250 251 al. (2019). The quality of the collagen extracts was assessed based on the collagen yield as a 252 percentage of the original bone weight (minimum requirement 1%). The elemental and isotopic 253 ratios of the extracts (~0.5 mg) were measured at the MPI-EVA on a Thermo Finnigan Flash elemental analyser coupled to a Thermo Delta plus XP isotope ratio mass spectrometer (EA-254 255 IRMS). Where sufficient collagen was extracted, collagen was graphitised using the automated 256 graphitisation equipment (AGE) (Wacker, Němec, & Bourquin, 2010) in the Lab of Ion Beam 257 Physics at ETH-Zurich (Switzerland) and dated on a MIni CArbon DAting System 258 (MICADAS) accelerator mass spectrometer (AMS) (Wacker, Bonani, et al., 2010). Where the 259 extracted collagen yield was insufficient for graphitization, it was combusted to CO<sub>2</sub> and 260 measured directly using the gas interface system coupled to the gas ion source of the MICADAS 261 (Wacker et al., 2013) following the protocol described in Fewlass et al. (2019) (see Appendix 262 A1.9 for more details).

To improve stratigraphic information available for the sites from which the analysed specimens originated we also obtained radiocarbon dates from five palaeontological sites (Appendix A1.10, Table S10). Radiocarbon dates were calibrated in OxCal v4.4 (Bronk Ramsey, 2009) using the IntCal20 (Reimer et al., 2020) calibration curve.

267

#### 268 **5 Results**

269 We generated a dataset of 4.2 kb long mtDNA sequences from 199 ancient and modern 270 common vole specimens (148 ancient and 51 modern). Sequences of 82 ancient specimens are 271 reported for the first time and either 4.2 kb or 1kb fragments of mtDNA of the remaining 66 272 specimens were reported previously (Baca et al., 2020; Baca et al., 2021; Lemanik et al., 2020) 273 (Table S1). All 82 newly reported specimens yielded short inserts and elevated level of cytosine 274 deamination at the terminal nucleotides characteristic for ancient DNA (Table S1). Ancient 275 specimens which yielded mtDNA sequences came from 40 sites scattered across Europe and 276 covered the period of the last ca. 60 thousand years (Table S1, Figure 1a). MtDNA cytochrome 277 b sequences of most modern specimens were reported previously (García et al., 2020; Stojak et 278 al., 2016) and here only the longer mtDNA fragment was generated to increase the phylogenetic 279 resolution. Direct radiocarbon dating was undertaken for 12 vole mandibles and 10 yielded 280 collagen of sufficient quality for AMS dating (Table S3). The carbon to nitrogen ratio (C:N) of 281 one specimen from Geißenklösterle (MI935) was at the limit (3.6) of accepted values for well-282 preserved collagen (2.9-3.6; Van Klinken, 1999) and yielded a relatively recent date with 283 respect to the latest Electron Spin Resonance dating of the cave sediments (Richard et al., 2019) 284 indicating that contamination with modern carbon, and therefore under-estimation of the true 285 age, is likely. We therefore discarded this radiocarbon date and estimated the age of this 286 specimen using the molecular approach. All other direct dates are regarded as reliable based on 287 their chemical indicators (collagen yield, stable isotopic and elemental values; Table S3). As a 288 result, we used sequences of 20 directly radiocarbon dated and 51 extant specimens to calibrate 289 the molecular clock. The analysis of temporal signal (BETS) showed that the dated specimens 290 included in our dataset are suitable for calibration of the molecular clock. The heterochronous 291 model, with a strict clock and correct sampling times assigned to specimens, was strongly 292 supported over all other models  $(2\ln BF > 9)$  (Table S4). The leave-one-out analysis revealed 293 that the dated dataset enables relatively accurate estimation of specimen ages, although in the 294 case of three specimens (MI074, MI1337 and MI1355) the 95% highest posterior density 295 intervals (95% HPD) of estimated ages did not overlap within 2-sigma ranges of the 296 radiocarbon dates (Figure S1; Appendix A). In addition, the estimated ages of most specimens 297 agreed with their stratigraphic position, providing evidence for accuracy of this approach (Table 298 S1).

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- 300



301

302 Figure 1. Sampling localities of modern and ancient common voles across Europe (a). Filled symbols 303 represent paleontological sites, while unfilled circles denote localities of modern specimens. Numbers 304 near modern localities correspond to specimen numbers (WM...) in Figure 2 and Table S2. Symbols 305 and names are consistent with those in Figure 2 and are coloured according to the vole mtDNA lineage 306 found at the site. If multiple mtDNA lineages were found at the site, two or more symbols are presented. 307 Distribution of the main mtDNA lineages in extant populations (b). The grey area depicts the current 308 range of common vole in Europe. The coloured circles represent sampling localities of the common vole 309 and mtDNA lineages compiled from previous studies: pink - Eastern (E); yellow - Central (CEN); 310 orange -- Italian (ITA); green -- Western-North (WN); violet -- Western-South (WS); navy blue -- Balkan 311 (B).

#### 312 5.1 Diversification of common vole mtDNA lineages

The maximum clade credibility tree obtained in BEAST 1.10.4 (Figure 2), recovered, with high support, all six mtDNA lineages characterised previously in analyses of the mtDNA cytochrome *b* of modern individuals (Bužan et al., 2010; Haynes et al., 2003; Heckel et al., 2005). In addition, we identified three lineages which were only present in the Late Pleistocene specimens. These lineages were named by their geographic provenance and phylogenetic position: WNII and WNIII in Western Europe (Germany and France) and ITAII in Italy and Croatia (Figure 1a and 2).



320

Figure 2. Maximum clade credibility tree of common vole mtDNA obtained in BEAST 1.10.4 and calibrated with radiocarbon dated specimens. Black dots indicate nodes with posterior probability above 0.95 and grey bars show the 95% HPD intervals of node ages. The tips are annotated with sample names, colour symbols (consistent with those in Figure 1) and, medians of age estimates (black font) or medians of calibrated radiocarbon dates (red font). The empty circles represent modern specimens. The legend

on the right side of the figure provides information on stratigraphy and dating of the source sites and
layers. The green and blue strips indicate the main interstadials and stadials: B – Brørup; H–C –
Hengelo–Charbon; D–G Denekamp–Grand Bois; LGM – Last Glacial Maximum, B–A – Bølling–
Allerød; YD – Younger Dryas.

330

The tMRCA of all European common voles was estimated to 90 ka ago (95% HPD: 98– 83 ka ago). Divergence time estimates of the subsequent lineages ranged between 82 ka ago (95% HPD: 91–75 ka ago) for the split between B and ITA/C/E, through 77 ka ago (95% HPD: 84–70 ka ago) for the split between WNIII and WNII/WN/WS to ca. 50–45 ka ago for the splits between a number of lineages: ITA and ITAII; CEN and E; WNII and WN (Figure 2).

#### **5.2** Temporal population structure and dynamics of the common vole

#### 337 5.2.1 Western and Southwestern Europe

338 The oldest specimens from Western Europe were classified as belonging to the WNIII 339 lineage which was the sister to the WS and WN/WNII lineages. The WNIII lineage contains 340 mainly individuals from the lowermost layers of Geißenklösterle (GH23-18). The age of these 341 individuals was estimated to between 56 and 45 ka (Figure 1a, 2, Table S1). Three haplotypes 342 of similar age as the latter (57–44 ka ago) from Western France (Roc-en-Pail; MI122, MI123) 343 and Northern Spain (El Portalon P9; MI1279) are located at the base of the WS, WNII and WN 344 lineages (pre-WN/WS; Figure 1a, Table S1). Around 42 ka ago the WS lineage appeared in 345 Northern Spain and the WNII lineage appeared in France, Belgium and Germany. The latter, 346 composed of specimens from the younger layers of Geißenklösterle, Trou Al'Wesse and 347 Jovelle, disappeared around 32 ka ago. The age of the oldest specimens from the WN lineage, 348 which come from the Trou Al'Wesse, was estimated to 37 ka ago. This lineage was found 349 among Late Pleistocene specimens from Western Germany, Belgium, France, and the UK.

350 The record from Spain suggests population continuity throughout the last 40 ka, 351 although nearly all modern and Holocene specimens coalesce around 11.5 ka ago (Figure 2), which suggests significant reduction of population size around the Pleistocene/Holocene 352 353 transition. This was further confirmed by the Bayesian demographic analysis, which suggest 354 about five-fold reduction of female effective population size around the Pleistocene/Holocene 355 transition with the minimum around the Early Holocene (11.7–9 ka ago), followed by a slight 356 recovery around the Middle Holocene (9–6 ka ago) (Appendix A1.8, Figures S2 and S3). The 357 WS lineage was also detected in southern France at Jovelle and Coulet des Roches during MIS 358 2 (29–14 ka ago).

#### 359 5.2.2 Central and Southeastern Europe

360 The oldest specimens in Central and Southeastern Europe come from Obłazowa 2, Nový 361 and layer 12 of Peskö, all from the Western Carpathians. Their ages were estimated to between 362 55 and 35 ka ago and they hold a basal position with respect to Central and Eastern lineages 363 (pre-CEN/E). The two oldest specimens from Peskö (MI299, MI300) were noticeably more 364 divergent than the others. Starting from 35 ka ago we found the CEN lineage in the Western 365 Carpathians as well as to their north, and the most recent specimens belonging to this lineage 366 were dated to the Early Holocene. The earliest specimens assigned to the Eastern lineage were 367 estimated to ca. 27–25 ka ago and were found in Central Romania (Muierilor Cave; MI760) 368 and Northwestern Bulgaria (Cave 16; MI807). The more recent specimens from this lineage 369 seem to represent a north and northwestern expansion of this population, with the first appearance in the Ukrainian Carpathians in Perlyna at around 14 ka ago and in the Western 370 371 Carpathians around 12 ka ago (Rejtek III, Muráň 3/1).

The Balkan lineage occupied the same area as its current distribution starting from at least 50 ka ago (Mujina pećina). A single specimen from Bivak Cave in the Northern Hungary suggests that prior to the Holocene, its range extended further to the northeast.

The ITA and ITAII lineages were detected on the Italian and Balkan Peninsulas. ITAII specimens were found in central Italy (Grotta del Sambuco) and in middle Dalmatia (Mujina pećina) with ages estimated to between 36.5 and 18.5 ka ago. These were generally older than specimens from the ITA lineage, dated to 22.6–0 ka ago and found in northern Italy (Riparo Tagliente) and Istria (Ljubićeva pećina). The single individual from layer 12 of Peskö cave (Western Carpathians; MI1701), directly radiocarbon dated to 45 ka cal BP, was located at the base of the ITA lineages.

#### 382 6 Discussion

383 The mitochondrial diversity of the common vole has been studied in detail to reconstruct the evolutionary history of species and elucidate its reactions to climate change. However, 384 385 inferences from modern mitochondrial diversity are limited, since the signal of the population 386 history is usually erased by the most recent reduction of effective population size (Mourier et 387 al., 2012). Ancient DNA enables direct observation of changes in genetic diversity in response to climate or environmental changes by sampling populations prior and after such changes. 388 389 Another great advantage of ancient DNA is that directly dated specimens may be used to 390 estimate substitution rates and to calibrate the molecular clock providing robust estimates of 391 divergence times without the need of external calibration. In this study, we make use of these

advantages and reconstructed the evolutionary history of the common vole at much greatertemporal depth providing new insights into its paleoecology.

394

# 6.1 The effects of climatic changes on diversification of common vole lineages

395 The estimated tMRCA of mtDNA for the European common vole (90 ka ago; 95% 396 HPD: 98–83 ka ago) is substantially older than some previous estimates (García et al., 2020; 397 Heckel et al., 2005; Stojak et al., 2015). It was similar to the recent estimates for the initial 398 diversification of mtDNA lineages of the cold-adapted collared lemmings (100 ka ago, 95% 399 HPD: 109–92 ka ago; E. Lord, personal communication). It was suggested that this may be an 400 effect of a bottleneck during the Eemian Interglacial (MIS5e). It may also be related to the 401 Brørup Interstadial (MIS 5c; ~GS-23; ca. 104–88 ka). Vegetation during the Brørup Interstadial 402 was characterised by temperate deciduous forests in Western Europe and boreal forests more 403 to the north and east (Guiter et al., 2003; Helmens, 2014), providing unfavourable habitats for 404 the cold-adapted collared lemming as well as for the common vole, both of which rely on 405 various types of open habitat.

406 The initial divergence of the common vole lineages may have been the result of survival 407 in two distinct refugial areas in the Alpine and Carpathian regions causing a partition in the 408 mitochondrial diversity of the species in with two geographical areas. Western Europe, 409 including the territories of present-day Spain, France, Switzerland, and parts of Germany, was 410 occupied by the WNIII, WNII, WN and WS lineages. Meanwhile, Central and South-eastern 411 Europe, including territories to the East and to the South of Germany, was occupied by the 412 CEN, ITAII, ITA, B and E lineages. This geographic partitioning was maintained for at least 413 45 ka (Figure 3) and probably contributed to patterns of restricted geneflow and partial reproductive isolation of the present-day populations of Western-North and Central lineages 414 415 (Beysard & Heckel, 2014).

416 The divergence of the main common vole mtDNA lineages estimated in this study was 417 also older than previously suggested. All mtDNA lineages present in extant European 418 populations (WS, WN, ITA, B, CEN, E) diverged prior to 40 ka ago, and the earliest specimens 419 belonging to each of those lineages in our dataset pre-date 25 ka ago. This suggests, contrary 420 to previous hypotheses (García et al., 2020; Heckel et al., 2005; Stojak et al., 2015), that climate 421 deterioration during the LGM did not play the major role in the initial divergence of the main 422 extant mtDNA lineages. However, the population decline and increased isolation during the 423 period of the most inhospitable climatic conditions may have reinforced the previously existing 424 divergence.

Most of the observed divergence events occurred between ca. 60 and 45 ka ago. They may be related to a long interstadial period identified in the palynological records across Europe: the Moershoofd Interstadial Complex in the Netherlands, the Pile Interstadial Complex at La Grande Pile pollen sequence (eastern France) and the Oerel and Glinde Interstadials at the Oerel pollen sequence (Northern Germany) dated to ca. 58–48 ka uncal BP and is usually correlated with Greenland Interstadials (GI) 16 and 14 (ca. 58–56.5 and 54–49.5 ka ago) (Helmens et al., 2014).

#### 432 **6.2 Phylogeography and demographic history of the common vole**

#### 433 6.2.1 Western Europe and Iberian Peninsula

434 In Western Europe, we documented two consecutive mtDNA lineage turnovers at the 435 end of MIS 3 (WNIII/WNII and WNII/WN). The first one took place around 45 ka ago (Figure 436 3). In our dataset there are few specimens of similar age from other parts of Europe; however, the divergent position of three specimens from the Western Carpathians (MI299, MI300, and 437 438 MI1701; Figure 2) dated to 53 and 45 ka ago suggests that synchronous lineage turnovers may 439 also have taken place in other parts of Europe. The second one occurred around 32 ka ago and 440 appears to have been restricted to Western Europe (Figure 3). The oxygen isotope record from 441 Greenland ice cores shows that the period between 45 and 29 ka ago was characterised by multiple short-term climatic oscillations (Rasmussen et al., 2014). Palynological records 442 443 revealed two main interstadials which stand out during this period and have been identified in 444 most of sediment sequences across Europe (Helmens, 2014). The older one, named Hengelo 445 from its type locality in the Netherlands or Charbon in the Grande Pile sequence from eastern 446 France, took place around 43-41 ka cal BP (38-36 ka uncal BP) (Helmens, 2014; 447 Vandenberghe & van der Plicht, 2016), whilst the younger one (Denekamp – Grand Bois) occurred around 34-33 ka cal BP (31-29 ka uncal BP) (Guiter et al., 2003; Helmens, 2014), 448 449 approximately at the same time as the recorded mtDNA lineage turnovers. The exact correlation 450 of palynological data with Greenland ice-core records is problematic due to potential offsets 451 and the wide error ranges involved. Hengelo-Charbon is usually associated with GI-11 (ca. 452 43.3–42.2 ka ago) or GI-10 (ca. 41.5–40.8 ka ago), although at times to the earlier GI-12 (ca. 453 46.8–44.2 ka ago) as the longest and most pronounced interstadial around the end of MIS 3, 454 and Denekamp–Grand Bois with GI-8 (38.2–36.6 ka ago) (Helmens, 2014). Both interstadials 455 were characterised by the emergence of Betula and Pinus forests in Western Europe, and of 456 Betula, Larix and, Pinus forests in Central Europe, although it is assumed that the landscape 457 remained relatively open as the duration of these interstadials was too short for the development

458 of full forest cover (Guiter et al., 2003; Helmens, 2014). It was shown, however, that even partitioned landscapes limit dispersal and promote local extinctions of common vole 459 460 populations (Delattre, Giraudoux, Baudry, Quere, & Fichet-Calvet, 1996), thus fragmentation 461 of primarily open, stadial habitats into patchy and mosaic interstadial landscapes might have 462 led to large scale decreases of population density and local or regional extinctions. A similar 463 explanation for common vole population dynamics was previously suggested by Tougard et al. 464 (2008). Martínková et al. (2013) showed partial replacement of mtDNA within the WN lineage 465 in Late Holocene common vole populations from northern France and Belgium, and also suggested that the main factor driving this process was likely landscape reorganization. 466

467 The record from Northern Spain, occupied by the WS lineage, suggests population 468 continuity throughout the last ca. 45 ka. The demographic reconstruction showed a drastic 469 reduction of the effective population size around the Pleistocene/Holocene transition (Figures 470 S2 and S3). This is in agreement with previous findings based on mtDNA cytochrome b and a 471 smaller sample size (Baca et al., 2020). Palynological records from the region from which both 472 modern and ancient specimens originated suggest that during the Bølling-Allerød interstadial 473 (14.7-12.8 ka ago) a high proportion of open landscapes persisted until the expansion of 474 deciduous woodlands started in the Early Holocene (Carrión et al., 2010).

475



476

Figure 3. Temporal distribution of the mtDNA lineages divided by the geographical regions. Western
Europe here includes France, southern Germany, Belgium, and the United Kingdom; the Northern

Europe includes northern Germany and Poland (there were no ancient specimens from this region), the Carpathians include southern Poland, Czechia, Slovakia, Hungary, Ukraine and Romania and the Balkans include Bulgaria, Serbia and Croatia; Circles denote medians of age estimated using molecular approach while squares denote medians of calibrated radiocarbon ages. The green and blue strips indicate the main interstadials and stadials as in Figure 2.

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- 485

## 6.2.2 Central and South-eastern Europe

486 The survival of common vole populations throughout the LGM at high latitudes, including the Carpathians, has been previously suggested based on multiple lines of evidence. 487 488 The fossil record suggests the continuous presence of the common vole in the Pannonian Basin 489 (Pazonyi, 2004) and even north of the Carpathians (Sommer & Nadachowski, 2006), although 490 these findings were based only on the stratigraphic position of the specimens, rather than direct 491 dates. The Carpathians was also suggested as a northern refugium based on the distribution of 492 the Eastern mtDNA haplogroup in modern populations (Stojak et al., 2016, 2015) and 493 ecological niche modelling (Stojak, Borowik, Górny, McDevitt, & Wójcik, 2019). Our data 494 support a northern survival of the common vole throughout the LGM. In the Western 495 Carpathians, the northernmost part of this mountain range, we found all individuals with ages 496 estimated, or directly dated, to between 36 and 10 ka ago to belong to the CEN mtDNA lineage. 497 Two specimens from Šarkanica yielded pre-LGM, direct radiocarbon dates (28.9 and 27.8 ka 498 cal BP; Table S3) while the age of a further four specimens was estimated to between 25.9 and 499 24.2 ka ago (Figure 2, Table S1). Recently, Lemanik et al. (2020) found a signal of rapid growth 500 of the effective female population size (Ne<sub>f</sub>) of the common vole population from the Western 501 Carpathians starting ca. 21 ka cal BP that continued until ca. 15 ka cal BP. Together, these 502 results are consistent with population continuity through the LGM, although accompanied by a 503 significant reduction of population size.

504 Most of Central, Eastern, and Southeastern Europe is at present occupied by the Eastern 505 lineage. It was suggested, based on both modern and ancient DNA, that the expansion of this 506 lineage started around the Pleistocene/Holocene transition (Baca et al., 2020; Stojak et al., 507 2016). Our new data allow us to refine the history of this lineage. Prior to the maximal extension 508 of the Scandinavian Ice Sheet (23-19 ka ago), we detected the Eastern lineage only in 509 Southwestern Romania and Bulgaria. The presence of younger specimens bearing this mtDNA 510 lineage in the same area, dated to 18.8 and 14 ka ago, suggests that this lineage may have 511 survived the LGM in this part of the Carpathians. The expansion of the Eastern lineage from

512 the South-eastern Carpathian area is also consistent with the finding of the highest genetic 513 diversity in the extant common vole populations in this area (Stojak et al., 2016). The 514 colonization of vast territories of Central and Eastern Europe, and replacement of Central and 515 Balkan mtDNA lineages, took place somewhat later during the Bølling–Allerød or the Younger 516 Dryas. In contrast to previous suggestions (Baca et al., 2020), the similar ages of Central and 517 Eastern lineage individuals from the Muráň 3 and Býčí sites estimated here, suggest that both 518 lineages may have coexisted in the area for some time (Figure 2, Table S1), prior to the final 519 extirpation of the Central lineage in the Early Holocene. Although, the limited resolution of 520 molecular age estimation and lack of possibility to track admixture with mtDNA does not allow 521 for fine scale reconstruction of this process.

522 The record from the Italian peninsula suggests a replacement of the ITAII lineage with 523 the ITA lineage which survived in Northern Italy and Switzerland until the present day. 524 Although the available data come from sites with a limited temporal span (Appendix A), our 525 dated phylogeny suggests that the extirpation of the ITAII lineage took place at some point after 526 the LGM and the expansion of the ITA lineage occurred no later than the Bølling-Allerød 527 warming. This lineage turnover may be reflected in the significant decrease in common vole 528 remains for a short period after the LGM which was observed in the southern Italian peninsula 529 (Berto, López-García, & Luzi, 2019). This is more consistent with repeated southward 530 expansions of subsequent common vole populations rather than with their continuous presence 531 in the region.

## 532 6.2.3 Comparison with other European Late Pleistocene species

533 The common vole, along with the collared lemming and narrow-headed voles, are 534 amongst the most numerous small mammals found in the assemblages from the last glacial 535 period in Europe and are assumed to have coexisted during much of this period. The explanation 536 for this paradox, in which a temperate species coexisted with cold-adapted ones, is the high 537 tolerance of common vole to low temperatures (Tougard et al., 2008). Our study corroborates 538 the continuous presence of the species at middle and high latitudes of Europe throughout the 539 last at least 60 ka (Figure 3), although the evolutionary history of the common vole differed in 540 some respects from other species inhabiting Europe during the Late Pleistocene.

The long-term regional continuity of the main lineages of common vole with limited evidence for migrations suggested by our mtDNA analysis is in contrast with the Late Pleistocene evolutionary histories of megafaunal species such as cave bears and mammoths, both of which showed evidence for long-distance migrations and large-scale population replacements

545 (Fellows Yates et al., 2017; Gretzinger et al., 2019). The collared lemming also showed a distinct pattern of mtDNA diversity consistent with multiple continent-wide population 546 547 replacements (Palkopoulou et al., 2016). This was likely related to differences in mobility of 548 the two species, with collared lemmings being more capable of long-distance dispersals than 549 the common vole (Ehrich et al., 2001). On the other hand, available data suggest that despite 550 this apparently different, species-specific history, common vole populations were affected by 551 the same climatic and environmental changes as the cold adapted taxa. The extirpation of the 552 WNIII lineage in Western Europe, and potentially of other common vole populations across 553 Europe, took place around the same time as the continent-wide disappearance of collared 554 lemming populations represented by their mtDNA lineages 1 and 2 (Palkopoulou et al., 2016). 555 Similarly, the disappearance of the WNII lineage of the common voles around 32 ka ago is at 556 approximately the same time as the population replacement of cave bears recorded in the Ach 557 Valley, Germany (Münzel et al., 2011), and of woolly mammoth populations across the whole 558 of Europe (Fellows Yates et al., 2017; Palkopoulou et al., 2013). Around 32 ka ago a new 559 population of collared lemming (mtDNA lineage 3) appeared in Europe after a potential short-560 term extirpation (Palkopoulou et al., 2016). It was suggested that the main driver affecting 561 mammalian species throughout the last glaciation were the abrupt warmings occurring at the 562 onset of the Greenland Interstadials (Cooper et al., 2015). Nevertheless, whether it was the 563 abruptness of the climatic changes, or the subsequent emergence of interstadial environments 564 is unclear, although the high environmental instability in the period between 45 and 29 ka ago 565 appears to have influenced a whole spectrum of differently adapted species including the 566 common vole.

567 The other period that seems to have had a large impact on common vole populations 568 was the Pleistocene/Holocene transition. The climatic warming and the emergence of forests 569 during the Bølling–Allerød interstadial (14.7–12.8 ka ago) and in the Early Holocene (11.7–9 570 ka ago) are considered a main causes of extirpation of many cold-adapted species from Europe 571 including the collared lemming and narrow-headed vole (Berto, Szymanek, et al., 2021; Royer 572 et al., 2016). Our data confirmed previous findings indicating a substantial population decline 573 of the common vole in Northern Spain, and mtDNA lineage replacement in the Western 574 Carpathians (Baca et al., 2020). In addition, we identified a potential lineage turnover on the 575 Italian peninsula. This, together with a probable extirpation of the common vole from the 576 British Isles in the Early Holocene (Baca et al., 2020), suggests a continent-wide impact of 577 interstadial environments on common vole populations.

#### 578 7 Conclusions

579 Our study shows that the initial diversification of Last Glacial and extant common vole 580 took place around 90 ka ago, during the Brørup interstadial (MIS 5c, GI-23). The divergence 581 of mtDNA lineages present in extant common vole populations, as well as the first appearance 582 of specimens belonging to those lineages, occurred earlier than previously estimated, mostly 583 during the MIS 3 (57–29 ka ago). At high latitudes of Europe, we detected lineage turnovers 584 whose dating suggest that they were likely caused by the fragmentation of primary habitats 585 through reforestation during the interstadials occurring between 45 and 29 ka ago. In 586 comparison, the climate deterioration during the LGM appears to have had a milder effect on 587 common vole populations. More recent demographic changes and lineage turnovers like those 588 recorded in Spain, the Western Carpathians, and Italy took place after the LGM or around the 589 Pleistocene/Holocene transition and were also likely related to climatic warming. Altogether, 590 this suggests that during the last glacial period, the evolutionary history of the common vole 591 was distinct from typical cold-adapted species associated with steppe-tundra environments, 592 although they responded to similar climatic and environmental changes. However, in contrast 593 to the collared lemming and narrow-headed vole, the common vole did not become extinct in 594 Europe at the end of the Pleistocene, possibly due to higher ecological plasticity, and eventually 595 expanded during the mid-Holocene, taking advantage of secondary habitats such as agricultural 596 fields. Overall, this suggests that habitat availability, rather than climatic variables, is the 597 primary factor affecting common vole populations.

598 8 Competing interests

599 We have no competing interests.

## 600 9 References

- Baca, M., Nadachowski, A., Lipecki, G., Mackiewicz, P., Marciszak, A., Popović, D., ...
  Wojtal, P. (2017). Impact of climatic changes in the Late Pleistocene on migrations and
  extinction of mammals in Europe: four case studies. *Geological Quarterly*, *61*(2), 291–
  304. doi: 10.7306/gq.1319
- 605 Baca, M., Popović, D., Baca, K., Lemanik, A., Doan, K., Horáček, I., ... Nadachowski, A.
- 606 (2020). Diverse responses of common vole (*Microtus arvalis*) populations to Late Glacial
- and Early Holocene climate changes Evidence from ancient DNA. *Quaternary Science Reviews*, 233, 106239. doi: 10.1016/j.quascirev.2020.106239
- 609 Baca, M., Popović, D., Lemanik, A., Baca, K., Horáček, I., & Nadachowski, A. (2019). Highly

- divergent lineage of narrow-headed vole from the Late Pleistocene Europe. *Scientific Reports*, 9(1), 17799. doi: 10.1038/s41598-019-53937-1
- Baca, M., Popović, D., Lemanik, A., Fewlass, H., Talamo, S., Zima, J., Ridush, B., Popov, V.,
- 613 & Nadachowski, A. (2021) The Tien Shan vole (*Microtus ilaeus*; Rodentia: Cricetidae)
- as a new species in the Late Pleistocene of Europe. *Ecology and Evolution*, 11, 16113–
  16125.
- 616 Berto, C., López-García, J. M., & Luzi, E. (2019). Changes in the Late Pleistocene small-
- 617 mammal distribution in the Italian Peninsula. *Quaternary Science Reviews*, 225, 106019.
  618 doi: 10.1016/j.quascirev.2019.106019
- Berto, C., Nadachowski, A., Pereswiet-Soltan, A., Lemanik, A., & Kot, M. (2021). The Middle
  Pleistocene small mammals from the lower layers of Tunel Wielki Cave (KrakówCzęstochowa Upland): An Early Toringian assemblage in Poland. *Quaternary International*, 577, 52–70. doi: 10.1016/j.quaint.2020.10.023
- Berto, C., Szymanek, M., Blain, H.-A., Pereswiet-Soltan, A., Krajcarz, M., & Kot, M. (in press).
  Small vertebrate and mollusc community response to the latest Pleistocene-Holocene
  environment and climate changes in the Kraków-Częstochowa Upland (Poland, Central
  Europe). *Quaternary International*, doi: 10.1016/j.quaint.2021.09.010
- Beysard, M., & Heckel, G. (2014). Structure and dynamics of hybrid zones at different stages
  of speciation in the common vole (*Microtus arvalis*). *Molecular Ecology*, 23(3), 673–687.
  doi: 10.1111/mec.12613
- Braaker, S., & Heckel, G. (2009). Transalpine colonisation and partial phylogeographic erosion
  by dispersal in the common vole (*Microtus arvalis*). *Molecular Ecology*, *18*(11), 2528–
  2531. doi: 10.1111/j.1365-294X.2009.04189.x
- Bronk Ramsey, C. (2009). Bayesian Analysis of Radiocarbon Dates. *Radiocarbon*, 51(1), 337–
   360. doi: 10.1017/S0033822200033865
- Bužan, E. V., Förster, D. W., Searle, J. B., & Kryštufek, B. (2010). A new cytochrome *b*phylogroup of the common vole (*Microtus arvalis*) endemic to the Balkans and its
  implications for the evolutionary history of the species. *Biological Journal of the Linnean Society*, 100(4), 788–796. doi: 10.1111/j.1095-8312.2010.01451.x
- 639 Carrión, J. S., Fernández, S., González-Sampériz, P., López-Merino, L., Carrión-Marco, Y.,
- 640 Gil-Romera, G., ... Burjachs, F. (2010). Expected trends and surprises in the Lateglacial
- and Holocene vegetation history of the Iberian Peninsula and Balearic Islands. *Review of*
- 642 *Palaeobotany and Palynology*, *162*(3), 458–475. doi: 10.1016/j.revpalbo.2009.12.007
- 643 Chaline, J. (1972). Les rongeures du pléistocène moyen et supérieur de France:(systématique,

644 biostratigraphie, paléoclimatologie). In *Cahiers Paléontologie*. Paris: CNRS.

- 645 Cooper, A., Turney, C., Hughen, K. A., Barry, W., McDonald, H. G., & Bradshaw, C. J. A.
- 646 (2015). Abrupt warming events drove Late Pleistocene Holarctic megafaunal turnover.
  647 Science, 349, 1–8. doi: 10.1126/science.aac4315
- Dabney, J., Knapp, M., Glocke, I., Gansauge, M.-T., Weihmann, A., Nickel, B., ... Meyer, M.
- 649 (2013). Complete mitochondrial genome sequence of a Middle Pleistocene cave bear
- 650 reconstructed from ultrashort DNA fragments. *Proceedings of the National Academy of*
- 651 Sciences of the United States of America, 110(39), 15758–15763. doi: 652 10.1073/pnas.1314445110
- Darriba, D., Taboada, G. L., Doallo, R., & Posada, D. (2012). jModelTest 2: more models, new
  heuristics and parallel computing. *Nature Methods*, 9(8), 772–772. doi:
  10.1038/nmeth.2109
- Delattre, P., Giraudoux, P., Baudry, J., Quere, J. P., & Fichet-Calvet, E. (1996). Effect of
  landscape structure on Common Vole (*Microtus arvalis*) distribution and abundance at
  several space scales. *Landscape Ecology*, 288(5), 279–288.
- Duchene, S., Lemey, P., Stadler, T., Ho, S. Y. W., Duchene, D. A., Dhanasekaran, V., & Baele,
  G. (2020). Bayesian evaluation of temporal signal in measurably evolving populations. *Molecular Biology and Evolution*, 37(11), 3363–3379. doi: 10.1093/molbev/msaa163
- Ehrich, D., Jorde, P. E., Krebs, C. J., Kenney, A. J., Stacy, J. E., & Stenseth, N. C. (2001).
  Spatial structure of lemming populations (*Dicrostonyx groenlandicus*) fluctuating in
- 664 density. *Molecular Ecology*, 10(2), 481–495. doi: 10.1046/j.1365-294X.2001.01229.x
- 665 Fellows Yates, J. A., Drucker, D. G., Reiter, E., Heumos, S., Welker, F., Münzel, S. C., ...
- Krause, J. (2017). Central European Woolly Mammoth population dynamics: insights
  from Late Pleistocene mitochondrial genomes. *Scientific Reports*, 7(1), 17714. doi:
  10.1038/s41598-017-17723-1
- Fewlass, H., Tuna, T., Fagault, Y., Hublin, J. J., Kromer, B., Bard, E., & Talamo, S. (2019).
  Pretreatment and gaseous radiocarbon dating of 40–100 mg archaeological bone. *Scientific*
- 671 *Reports*, 9(1), 1–11. doi: 10.1038/s41598-019-41557-8
- Fink, S., Excoffier, L., & Heckel, G. (2004). Mitochondrial gene diversity in the common vole
   *Microtus arvalis* shaped by historical divergence and local adaptations. *Molecular Ecology*, 13, 3501–3514. doi: 10.1111/j.1365-294X.2004.02351.x
- Fischer, M. C., Foll, M., Heckel, G., & Excoffier, L. (2014). Continental-scale footprint of
  balancing and positive selection in a small rodent (*Microtus arvalis*). *PLoS ONE*, 9(11),
- 677 e112332. doi: 10.1371/journal.pone.0112332

- Folkertsma, R., Westbury, M. V., Eccard, J.A., & Hofreiter, M. (2018) The complete
  mitochondrial genome of the common vole, Microtus arvalis (Rodentia: Arvicolinae).
  Mitochondrial DNA Part B: Resources, 3, 446–447.
- Gansauge, M. T., Aximu-Petri, A., Nagel, S., & Meyer, M. (2020). Manual and automated
  preparation of single-stranded DNA libraries for the sequencing of DNA from ancient
  biological remains and other sources of highly degraded DNA. *Nature Protocols*, 15(8),
  2279–2300. doi: 10.1038/s41596-020-0338-0
- 227)-2300. doi: 10.1030/341370-020-0330-0
- 685 García, J. T., Domínguez-Villaseñor, J., Alda, F., Calero-Riestra, M., Pérez Olea, P., Fargallo,
- J. A., ... Viñuela, J. (2020). A complex scenario of glacial survival in Mediterranean and
  continental refugia of a temperate continental vole species (*Microtus arvalis*) in Europe. *Journal of Zoological Systematics and Evolutionary Research*, 58(1), 459–474. doi:
  10.1111/jzs.12323
- Gretzinger, J., Molak, M., Reiter, E., Pfrengle, S., Urban, C., Neukamm, J., ... Schuenemann,
  V. J. (2019). Large-scale mitogenomic analysis of the phylogeography of the Late
  Pleistocene cave bear. *Scientific Reports*, 9(1), 1–11. doi: 10.1038/s41598-019-47073-z
- Guiter, F., Andrieu-Ponel, V., de Beaulieu, J.-L., Cheddadi, R., Calvez, M., Ponel, P., ...
  Goeury, C. (2003). The last climatic cycles in Western Europe : a comparison between
  long continuous lacustrine sequences from France and other terrestrial records. *Quaternary International*, 111, 59–74. doi: 10.1016/S1040-6182(03)00015-6
- Haynes, S., Jaarola, M., & Searle, J. B. (2003). Phylogeography of the common vole (*Microtus arvalis*) with particular emphasis on the colonization of the Orkney archipelago. *Molecular Ecology*, 12(4), 951–956. doi: 10.1046/j.1365-294X.2003.01795.x
- Heckel, G., Burri, R., Fink, S., Desmet, J.-F., & Excoffier, L. (2005). Genetic structure and
  colonization processes in European populations of the common vole, *Microtus arvalis*. *Evolution; International Journal of Organic Evolution*, 59(10), 2231–2242.. doi:
  10.1554/05-255.1.
- Helmens, K. F. (2014). The Last Interglacial-Glacial cycle (MIS 5-2) re-examined based on
  long proxy records from central and northern Europe. *Quaternary Science Reviews*, 86,
  115–123. doi: 10.1016/j.quascirev.2013.12.012
- Horáček, I., & Ložek, V. (1988). Palaeozoology and the Mid-European Quaternary Past:
  Scope of the Approach and Selected Results. Praha: Rozpravy ČSAV, Řada MPV.
- Jacob, J., Manson, P., Barfknecht, R., & Fredricks, T. (2014). Common vole (*Microtus arvalis*)
- ecology and management: Implications for risk assessment of plant protection products.
- 711 Pest Management Science, Vol. 70, pp. 869-878. John Wiley & Sons, Ltd. doi:

- 712 10.1002/ps.3695
- Jánossy, D. (1986). Pleistocene vertebrate faunas of Hungary. In *Pleistocene vertebrate faunas of Hungary*. Amsterdam-Oxford-New York-Takyo: Elsevier. doi: 10.1016/00472484(89)90045-6
- Jónsson, H., Ginolhac, A., Schubert, M., Johnson, P. L. F., & Orlando, L. (2013).
  MapDamage2.0: Fast approximate Bayesian estimates of ancient DNA damage

parameters. *Bioinformatics*, 29(13), 1682–1684. doi: 10.1093/bioinformatics/btt193

- Katoh, K., & Standley, D. M. (2013). MAFFT multiple sequence alignment software version
  7: Improvements in performance and usability. *Molecular Biology and Evolution*, *30*(4),
  72. 772–780. doi: 10.1093/molbev/mst010
- Kučera, J., Suvova, Z., & Horáček, I. (2009). Early Middle Pleistocene glacial community of
  rodents (Rodentia): Stránská skála cave (Czech Republic). *Lynx*, (40), 43–69.
- 724 Lemanik, A., Baca, M., Wertz, K., Socha, P., Popović, D., Tomek, T., ... Nadachowski, A.
- (2020). The impact of major warming at 14.7 ka on environmental changes and activity of
  Final Palaeolithic hunters at a local scale (Orawa-Nowy Targ Basin, Western Carpathians,
  Poland). *Archaeological and Anthropological Sciences*, *12*(3). doi: 10.1007/s12520-02001020-6
- Li, H., & Durbin, R. (2010). Fast and accurate long-read alignment with Burrows-Wheeler
  transform. *Bioinformatics (Oxford, England)*, 26(5), 589–595. doi:
  10.1093/bioinformatics/btp698
- Li, H., Handsaker, B., Wysoker, A., Fennell, T., Ruan, J., Homer, N., ... Subgroup, 1000
  Genome Project Data Processing. (2009). The Sequence Alignment/Map format and
  SAMtools. *Bioinformatics*, 25(16), 2078–2079. doi: 10.1093/bioinformatics/btp352
- Lischer, H. E. L., Excoffier, L., & Heckel, G. (2014). Ignoring heterozygous sites biases
  phylogenomic estimates of divergence times: Implications for the evolutionary history of *Microtus* voles. *Molecular Biology and Evolution*, 31(4), 817–831. doi:
  10.1093/molbev/mst271
- Lorenzen, E. D., Nogués-Bravo, D., Orlando, L., Weinstock, J., Binladen, J., Marske, K. A., ...
  Willerslev, E. (2011). Species-specific responses of Late Quaternary megafauna to climate
  and humans. *Nature*, 479(7373), 359–364. doi: 10.1038/nature10574
- 742 Martínková, N., Barnett, R., Cucchi, T., Struchen, R., Pascal, M., Pascal, M., ... Searle, J. B.
- (2013). Divergent evolutionary processes associated with colonization of offshore islands.
   *Molecular Ecology*, 22(20), 5205–5220. doi: 10.1111/mec.12462
- 745 Maul, L. C., & Markova, A. K. (2007). Similarity and regional differences in Quaternary

arvicolid evolution in Central and Eastern Europe. *Quaternary International*, 160(1), 81–

747 99. doi: 10.1016/j.quaint.2006.09.010

Meyer, M., & Kircher, M. (2010). Illumina sequencing library preparation for highly
multiplexed target capture and sequencing. *Cold Spring Harbor Protocols*, 5(6), t5448.

750 doi: 10.1101/pdb.prot5448

- Milne, I., Stephen, G., Bayer, M., Cock, P. J. A., Pritchard, L., Cardle, L., ... Marshall, D.
  (2013). Using Tablet for visual exploration of second-generation sequencing data. *Briefings in Bioinformatics*, 14(2), 193–202. doi: 10.1093/bib/bbs012
- Mourier, T., Ho, S.Y.W., Gilbert, M.T.P., Willerslev, E., & Orlando, L. (2012) Statistical
   guidelines for detecting past population shifts using ancient DNA. *Molecular Biology and Evolution*, 29, 2241–51. doi:10.1093/molbev/mss094
- Münzel, S. C., Stiller, M., Hofreiter, M., Mittnik, A., Conard, N. J., & Bocherens, H. (2011).
  Pleistocene bears in the Swabian Jura (Germany): Genetic replacement, ecological
  displacement, extinctions and survival. *Quaternary International*, 245(2), 1–13. doi:
  10.1016/j.quaint.2011.03.060
- Nadachowski, A. (1989). Origin and history of the present rodent fauna in Poland based on
  fossil evidence. *Acta Theriologica*, 34(2), 37–53.
- Palkopoulou, E., Baca, M., Abramson, N. I., Sablin, M., Socha, P., Nadachowski, A., ... Dalén,
  L. (2016). Synchronous genetic turnovers across Western Eurasia in Late Pleistocene
  collared lemmings. *Global Change Biology*, 22(5), 1710–1721. doi: 10.1111/gcb.13214

766 Palkopoulou, E., Dalen, L., Lister, A. M., Vartanyan, S., Sablin, M., Sher, A., ... Thomas, J. A.

- 767 (2013). Holarctic genetic structure and range dynamics in the woolly mammoth.
  768 Proceedings of the Royal Society B: Biological Sciences, 280, 20131910. doi:
  769 10.1098/rspb.2013.1910.
- Pazonyi, P. (2004). Mammalian ecosystem dynamics in the Carpathian Basin during the last
  27,000 years. *Palaeogeography, Palaeoclimatology, Palaeoecology, 212*(3–4), 295–314.
  doi: 10.1016/j.palaeo.2004.06.008
- Rasmussen, S. O., Bigler, M., Blockley, S. P., Blunier, T., Buchardt, S. L., Clausen, H. B., ...
  Winstrup, M. (2014). A stratigraphic framework for abrupt climatic changes during the
  Last Glacial period based on three synchronized Greenland ice-core records: refining and
- extending the INTIMATE event stratigraphy. *Quaternary Science Reviews*, *106*, 14–28.
  doi: 10.1016/j.quascirev.2014.09.007
- Reimer, P. J., Austin, W. E. N., Bard, E., Bayliss, A., Blackwell, P. G., Bronk Ramsey, C., ...
  Talamo, S. (2020). The IntCal20 northern hemisphere radiocarbon age calibration curve

780 (0–55 cal kBP). *Radiocarbon*, 62(4), 725–757. doi: 10.1017/rdc.2020.41

- 781 Richard, M., Falguères, C., Valladas, H., Ghaleb, B., Pons-Branchu, E., Mercier, N., ... Conard,
- 782 N. J. (2019). New electron spin resonance (ESR) ages from Geißenklösterle Cave: A
- chronological study of the Middle and early Upper Paleolithic layers. *Journal of Human Evolution*, 133, 133–145. doi: 10.1016/j.jhevol.2019.05.014
- Royer, A., Montuire, S., Legendre, S., Discamps, E., Jeannet, M., & Lécuyer, C. (2016).
  Investigating the influence of climate changes on rodent communities at a regional-scale
- 787 (MIS 1-3, Southwestern France). *PLoS ONE*, *11*(1), e0145600. doi:
  788 10.1371/journal.pone.0145600
- Schubert, M., Lindgreen, S., & Orlando, L. (2016). AdapterRemoval v2: Rapid adapter
  trimming, identification, and read merging. *BMC Research Notes*, 9(1), 1–7. doi:
  10.1186/s13104-016-1900-2
- Socha, P. (2014). Rodent palaeofaunas from Biśnik Cave (Kraków-Częstochowa Upland,
   Poland): Palaeoecological, palaeoclimatic and biostratigraphic reconstruction. *Quaternary International*, 326–327, 64–81. doi: 10.1016/j.quaint.2013.12.027
- Sommer, R. S., & Nadachowski, A. (2006). Glacial refugia of mammals in Europe: evidence
  from fossil records. *Mammal Review*, 36(4), 251–265. doi: 10.1111/j.13652907.2006.00093.x
- Stewart, J. R., Lister, A. M., Barnes, I., & Dalén, L. (2010). Refugia revisited: individualistic
  responses of species in space and time. *Proceedings. Biological Sciences / The Royal Society*, 277(1682), 661–671. doi: 10.1098/rspb.2009.1272
- Stojak, J., Borowik, T., Górny, M., McDevitt, A. D., & Wójcik, J. M. (2019). Climatic
  influences on the genetic structure and distribution of the common vole and field vole in
  Europe. *Mammal Research*, 64(1), 19–29. doi: 10.1007/s13364-018-0395-8
- Stojak, J., McDevitt, A. D., Herman, J. S., Kryštufek, B., Uhlíková, J., Purger, J. J., ... Wójcik,
  J. M. (2016). Between the Balkans and the Baltic: Phylogeography of a Common Vole
  mitochondrial DNA lineage limited to Central Europe. *PLOS ONE*, *11*(12), e0168621. doi:
- 807 10.1371/journal.pone.0168621
- Stojak, J., Mcdevitt, A. D., Herman, J. S., Searle, J. B., & Wójcik, J. M. (2015). Post-glacial
  colonization of eastern Europe from the Carpathian refugium: evidence from
  mitochondrial DNA of the common vole *Microtus arvalis*. *Biological Journal of the Linnean Society*, 115(4), 927–939.
- 812 Suchard, M. A., Lemey, P., Baele, G., Ayres, D. L., Drummond, A. J., & Rambaut, A. (2018).
- 813 Bayesian phylogenetic and phylodynamic data integration using BEAST 1.10. Virus

814 *Evolution*, 4(1), 1–5. doi: 10.1093/ve/vey016

- 815 Tougard, C., Renvoisé, E., Petitjean, A., & Quéré, J.-P. (2008). New insight into the
- 816 colonization processes of common voles: inferences from molecular and fossil evidence.

817 *PloS One*, *3*(10), e3532. doi: 10.1371/journal.pone.0003532

- 818 Van Klinken, G. J. (1999). Bone collagen quality indicators for palaeodietary and radiocarbon
- 819 measurements. Journal of Archaeological Science, 26(6), 687-695. doi:
- 820 10.1006/jasc.1998.0385
- Vandenberghe, J., & van der Plicht, J. (2016). The age of the Hengelo interstadial revisited. *Quaternary Geochronology*, *32*, 21–28. doi: 10.1016/j.quageo.2015.12.004
- 823 Wacker, L., Bonani, G., Friedrich, M., Hajdas, I., Kromer, B., Němec, M., ... Vockenhuber, C.
- 824 (2010). MICADAS: Routine and high-precision radiocarbon dating. *Radiocarbon*, 52(2),
   825 252–262. doi: 10.1017/S0033822200045288
- 826 Wacker, L., Fahrni, S. M., Hajdas, I., Molnar, M., Synal, H. A., Szidat, S., & Zhang, Y. L.
- 827 (2013). A versatile gas interface for routine radiocarbon analysis with a gas ion source.
- Nuclear Instruments and Methods in Physics Research, Section B: Beam Interactions with
  Materials and Atoms, 294, 315–319. doi: 10.1016/j.nimb.2012.02.009
- 830 Wacker, L., Němec, M., & Bourquin, J. (2010). A revolutionary graphitisation system: Fully
- automated, compact and simple. *Nuclear Instruments and Methods in Physics Research,*
- 832 Section B: Beam Interactions with Materials and Atoms, 268(7–8), 931–934. doi:
- 833 10.1016/j.nimb.2009.10.067
- Walther, G. R. (2010). Community and ecosystem responses to recent climate change. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 365(1549), 2019–
  2024. doi: 10.1098/rstb.2010.0021
- 837

## 838 **10 Data Availability**

The consensus mtDNA sequences generated in this study have been deposited in GenBank under accession numbers OL588336 - OL588524. The alignment used for the reconstruction of phylogeny have been deposited in Dryad (doi:10.5061/dryad.4j0zpc8d9). Mitochondrial alignments generated in this study have been deposited in the European Nucleotide Archive under project number PRJEB53474.

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## 845 11 Author Contributions

- 846 MB, DP and AN designed research; MB, DP, AL, HF and XW performed research; MB
- 847 analysed the data and prepared figures; SBC, NC, GCB, ED, JTG, GH, IH, MVK, LL, JMLG,
- 848 EL, ZM, JML, XM, AP, VP, TH, SER, BR, AR, JRS. JS, ST and JMW contributed samples;
- 849 MB and AN wrote the paper with the input from all co-authors.